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*Published in:*  
Applied and environmental microbiology

*DOI:*  
[10.1128/AEM.02327-09](https://doi.org/10.1128/AEM.02327-09)

**IMPORTANT NOTE:** You are advised to consult the publisher's version (publisher's PDF) if you wish to cite from it. Please check the document version below.

*Document Version*  
Publisher's PDF, also known as Version of record

*Publication date:*  
2010

[Link to publication in University of Groningen/UMCG research database](#)

### *Citation for published version (APA):*

Meijer, W. H., Gidijala, L., Fekken, S., Kiel, J. A. K. W., van den Berg, M. A., Lascaris, R., Bovenberg, R. A. L., & van der Klei, I. J. (2010). Peroxisomes are required for efficient penicillin biosynthesis in *Penicillium chrysogenum*. *Applied and environmental microbiology*, 76(17), 5702-5709.  
<https://doi.org/10.1128/AEM.02327-09>

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## Supplementary Tables

**Supplementary Table 1. Plasmids used in this study**

Name	Description	reference
pGBRH2	Plasmid containing $P_{pcbC}$ and $T_{penDE}$ , $Amp^R$	(2)
pFEM36	Plasmid containing GFP•SKL cassette, $Kan^R$	(4)
pGBRH2-GFP.SKL	pGBRH2 with GFP•SKL, $Amp^R$	This study
pBBK-001	pGBRH2 with DsRed•SKL, $Amp^R$	(1)
pNiGANi	Plasmid with <i>A. nidulans amdS</i> gene expressed by <i>A. nidulans gpdA</i> promoter, $Amp^R$	DSM, Lab collection
pBBK-007	pBBK-001 with $P_{pcbC}$ replaced by $P_{gpdA}$ , $Amp^R$	This study
pWHM-001	pGBRH2-GFP•SKL with $P_{pcbC}$ replaced by $P_{gpdA}$ , $Amp^R$	This study
pDONR221	Multisite Gateway vector, $Kan^R$ $Cm^R$	Invitrogen
pENTR221-pex3-nostop	pDONR221 with <i>P. chrysogenum pex3</i> gene lacking stop codon, $Kan^R$	This study
pDONR P4-P1R	Multisite Gateway vector, $Kan^R$ $Cm^R$	Invitrogen
pENTR41- $P_{gpdA}$	pDONR P4-P1R with <i>A. nidulans gpdA</i> promoter, $Kan^R$	Gift from J.G. Nijland
pDONR P2R-P3	Multisite Gateway vector, $Kan^R$ $Cm^R$	Invitrogen
pENTR23-GFP- $T_{penDE}$	pDONR P2R-P3 with <i>GFP</i> gene and <i>P. chrysogenum penDE</i> terminator, $Kan^R$	Gift from J.G. Nijland
pDEST R4-R3	Multisite Gateway vector, $Amp^R$ $Cm^R$	Invitrogen
pEXP-Pcpex3•GFP	pDEST R4-R3 with $P_{gpdA}$ - <i>pex3</i> •GFP- $T_{penDE}$ cassette, $Amp^R$	This study
pENTR221- $P_{gpdA}$ -GFP•SKL- $T_{penDE}$	pDONR221 with $P_{gpdA}$ -GFP•SKL- $T_{penDE}$ cassette from pWHM-001, $Kan^R$	This study
pENTR41-5'niaD	pDONR P4-P1R with 5'end of <i>P. chrysogenum niaD</i> gene; $Kan^R$	DSM, Lab collection
pENTR23-3'niaD	pDONR P2R-P3 with 3'end of <i>P. chrysogenum niaD</i> gene; $Kan^R$	DSM, Lab collection
pEXP-5'niaD- $P_{gpdA}$ -GFP•SKL- $T_{penDE}$ -3'niaD	pDEST R4-R3 with $\Delta niaD::P_{gpdA}$ -GFP•SKL- $T_{penDE}$ cassette, $Amp^R$	This study
pENTR41-DNM1-Prom	pDONR P4-P1R containing <i>P. chrysogenum dnm1</i> promoter region, $Kan^R$	This study
pENTR23-DNM1-Term	pDONR P2R-P3 containing <i>P. chrysogenum dnm1</i> terminator region, $Kan^R$	This study
pENTR221-AMDS	pDONR221 containing <i>A. nidulans amdS</i> expression cassette, $Kan^R$	Gift from J.G. Nijland.
pEXP43-5Dnm1-AMDS-3Dnm1	pDEST R4-R3 containing <i>P. chrysogenum dnm1</i> deletion cassette, $Amp^R$	This study
pDEST R4-R3/AMDS	pDEST R4-R3 with <i>A. nidulans amdS</i> expression cassette, $Amp^R$ $Cm^R$	(1)
pENTR41- $P_{pcbC}$	pDONR P4-P1R containing <i>P. chrysogenum pcbC</i>	(3)

	promoter region, Kan <sup>R</sup>	
pENTR221-Pcdnm1	pDONR221 containing <i>P. chrysogenum dnm1</i> cDNA, without stop codon, Amp <sup>R</sup>	This study
pENTR23-His8•T <sub>penDE</sub>	pDONR P2R-P3 containing His8 tag and <i>P. chrysogenum penDE</i> terminator region, Kan <sup>R</sup>	(3)
pEXP-PcDNM1•HIS8	Plasmid containing P <sub>pcbC</sub> - <i>dnm1</i> •His8-T <sub>penDE</sub> expression cassette, Amp <sup>R</sup>	This study

**Supplementary Table 2. Oligonucleotide primers used in this study**

<b>Name</b>	<b>Oligonucleotide primers</b>
BB-JK009	5' AGAGGTACCGCGGCCGCTCTGTACAGTGACCGGTGAC 3'
BB-JK010	5' AGAGGATCCTGTGATGTCTGCTCAAGCGGGG 3'
attB1-f-pex3	5` GGGGACAAGTTTGTACAAAAAAGCAGGCTCCATGATCGGTGCGACGAGGCG 3`
attB2-r-pex3-nostop	5` GGGGACCACTTTGTACAAGAAAGCTGGGTCCTGTGTTGGCTGCTTTTCTTC3`
attB1-f-PgpdA	5` GGGGACAAGTTTGTACAAAAAAGCAGGCTTAACCGCGGCCGCTCTGTACAGT 3`
attB2-r-TpenDE	5'GGGGACCACTTTGTACAAGAAAGCTGGGT ACTCCCCCCTGAAAGAGTTG 3'
BB-JK201	5'GGGGACAACCTTTGTATAGAAAAGTTGCTCGTTCGAGTTGCACCTTATCTG 3'
BB-JK202	5'GGGGACTGCTTTTTTGTACAAACTTGAGATCTTGAGGTTGATCGGCTGTC 3'
BB-JK203	5'GGGGACAGCTTTCTTGTACAAAGTGGCTGGTCTACGAGGAGCTTATCAAG
BB-JK204	5'GGGGACAACCTTTGTATAATAAAGTTGCGTCAAGCGTGACGCAGAATCAAG 3'
BB-JK205	5'TGTGGCCTATGTATCGTATCCTTG 3'
PgpdA.rev	5'GCATGCCAGAAAGAGTCAACC 3'
BB-JK209	5'GGGGACAAGTTTGTACAAAAAAGCAGGCTTTCGGTTCGACATCATGACATCGCTG 3'
BB-JK210	5'GGGGACCACTTTGTACAAGAAAGCTGGGTTTAACACCTCGCCACAATCTTGGC 3'
BB-JK211	5'TCTGCTCGTGACAGTGAACAAG 3'
BB-JK212	5'CTGCTTCGATCTCTTGCTTCAC 3'

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